

## VOLATILE CONSTITUENTS OF FLOWERS AND LEAVES OF *Anthemis hyalina*

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*The chemical composition of the essential oils of the flowers and leaves of Anthemis hyalina were analyzed by GC and GC-MS for the first time. The oils were found to contain seventy-two components. cis-Chrysanthenyl acetate (14.9% and 17.8%), camphor (11.6% and 1.7%), terpinen-4-ol (8.3% and 1.2%), germacrene-D (5.1% and 2.1%),  $\beta$ -caryophyllene (4.1% and 5.4%), myrcene (3.6% and 16.9%), bicyclogermacrene (3.5% and 0.9%),  $\alpha$ -pinene (2.3% and 4.1%), cis- $\beta$ -ocimene (2.1% and 4.3%) and isospathulenol (0.4% and 4.3%) were found to be the major constituents of the oils of flowers and leaves respectively.*

**Key words:** *Anthemis hyalina*, essential oil, GC/MS analysis, cis-chrysanthenyl acetate, camphor.

The genus *Anthemis* comprises more than 200 species and is considered one of the largest genera of the Compositae family [1]. The genus is represented in the flora of Iran by 39 species, including 15 endemics [2]. Several *Anthemis* species are used in Iranian folk medicine as medicinal plants [3]. The essential oil of *A. nobilis* possesses interesting anti-inflammatory and sedative properties in rat [4]. There are also reports on antimicrobial and larvicidal activities of the essential oils of *A. xylopoda* and *A. melampodina* respectively [5, 6]. The phytochemical studies of several *Anthemis* species have led to the isolation of sesquiterpene lactones, flavonoids, coumarins, acetylenic compounds, and essential oils [7–10].

*Anthemis hyalina* DC. is an Iranian native species known as “Babooneh Shafaf” in Persian [2]. According to the literature, *A. hyalina* has not been the subject of phytochemical research up to now. As part of a program of chemical investigation on the essential oil of Iranian aromatic plants, the chemical constituents of the essential oils from flowers and leaves of *A. hyalina* are reported for the first time.

The yield of essential oils obtained from flowers and leaves of the plant were 0.3% and 0.5% (v/w) respectively. Table 1 shows the composition of the essential oils obtained from flowers and leaves of *A. hyalina*. Compounds are listed in order of their elution from an HP-5 column. Seventy compounds of flower oil and sixty-four components of leave oil were identified. As can be seen, no great qualitative variations were observed in the composition of the flower and leaf essential oils; however, there are considerable quantitative variations in the percentages of some components of the oils. The major components of the oils of the flowers and leaves were cis-chrysanthenyl acetate (14.9% and 17.8%), camphor (11.6% and 1.7%), terpinen-4-ol (8.3% and 1.2%), germacrene-D (5.1% and 2.1%),  $\beta$ -caryophyllene (4.1% and 5.4%), myrcene (3.6% and 16.9%), bicyclogermacrene (3.5% and 0.9%),  $\alpha$ -pinene (2.3% and 4.1%), cis- $\beta$ -ocimene (2.1% and 4.3%), and isospathulenol (0.4% and 4.3%) respectively.

In spite of the large size of the genus *Anthemis*, the composition of volatile compounds is known in only a small number of species. The main constituents of the flower and leaf oils of *A. xylopoda*, *A. altissima*, and *A. altissima* (L.) var. *altissima* are reported as borneol (31.9% and 30.1%),  $\beta$ -caryophyllene (25.3% and 17.2%), and  $\beta$ -thujone (19.7% and 33.7%) respectively [5, 11–12]. Santolinatriene (27.3%), 1,8-cineole (7.9%), camphor (19.4%), and  $\alpha$ -thujone (40.2%) are reported as the major components of the essential oil of *A. melampodina*, *A. tinctoria*, *A. cretia*, and *A. carpatica* respectively [6, 13–15]. Chrysanthenyl acetate, the major component of the oils of *A. hyalina*, was also identified as the major fraction (11.3%) of the oil of *A. montana* [16].

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TABLE 1. Percentage Composition of the Essential Oils of *Anthemis hyalina* DC.

RI	Compound	Flower	Leaf	RI	Compound	Flower	Leaf
856	<i>trans</i> -2-Hexenal	0.1	0.2	1230	<i>cis</i> -Carveol	0.1	-
882	2-Methyl butyl acetate	0.1	1.9	1265	<b><i>cis</i>-Chrysanthenyl acetate</b>	<b>14.9</b>	<b>17.8</b>
927	Tricyclene	0.1	0.3	1279	<i>trans</i> -Carvone oxide	0.1	2.1
932	$\alpha$ -Thujene	0.2	Tr.	1288	Bornyl acetate	2.5	0.3
<b>940</b>	<b><math>\alpha</math>-Pinene*</b>	<b>2.3</b>	<b>4.1</b>	1294	<i>trans</i> -Sabinyl acetate	0.6	0.5
955	Camphene	1.9	0.3	1317	<i>trans, trans</i> -2,4-Decadienal	0.1	-
977	Sabinene	1.3	0.5	1328	<i>iso</i> -Dihydrocarveol acetate	1.7	1.4
981	$\beta$ -Pinene	1.1	1.0	1338	Bicycloelemene	1.6	0.3
993	<b>Myrcene*</b>	<b>3.6</b>	<b>16.9</b>	1373	$\alpha$ -Copaene	0.1	0.1
1003	$\delta$ -2-Carene	0.1	1.9	1381	$\beta$ -Bourbonene	0.2	Tr.
1007	$\alpha$ -Phellandrene	1.1	0.9	1388	$\beta$ -Elemene	0.4	0.2
1013	$\delta$ -3-Carene	-	0.6	1396	<i>n</i> -Tetradecane	0.2	-
1020	$\alpha$ -Terpinene	1.2	0.2	1416	<b><math>\beta</math>-Caryophyllene*</b>	<b>4.1</b>	<b>5.4</b>
1028	<i>p</i> -Cymene	0.7	0.8	1429	$\beta$ -Gurjunene	0.1	Tr.
1033	Limonene	0.1	0.2	1451	$\alpha$ -Humulene	1.1	1.6
1035	1,8-Cineole	3.1	0.3	1456	<i>trans</i> - $\beta$ -Farnesene	1.9	0.3
<b>1042</b>	<b><i>cis</i>-<math>\beta</math>-Ocimene</b>	<b>2.1</b>	<b>4.3</b>	1460	$\beta$ -Santalene	-	0.2
1046	Benzene acetaldehyde	0.1	-	1471	$\gamma$ -Gurjunene	0.2	0.3
1053	<i>trans</i> - $\beta$ -Ocimene	0.8	1.6	<b>1478</b>	<b>Germacrene D</b>	<b>5.1</b>	<b>2.1</b>
1065	$\gamma$ -Terpinene	2.3	0.6	1487	<i>cis</i> - $\beta$ -Guaiene	0.2	0.2
1071	<i>cis</i> -Sabinenehydrate	1.1	0.3	<b>1491</b>	<b>Bicyclogermacrene</b>	<b>3.5</b>	<b>0.9</b>
1091	Terpinolene	0.6	0.1	1506	( <i>E,E</i> )- $\alpha$ -Farnesene	0.5	0.1
1100	<i>trans</i> -Sabinenehydrate	0.2	0.8	1521	$\delta$ -Cadinene	0.2	0.1
1105	Nonanal	0.7	0.2	1562	<i>trans</i> -Nerolidol	0.3	0.8
1117	<i>trans</i> -Thujone	0.2	0.3	1574	Spathulenol	1.8	1.2
1128	$\alpha$ -Campholenal	0.3	0.1	1579	Caryophyllene oxide	0.9	2.5
1132	<i>allo</i> -Ocimene	0.2	0.3	1597	Guaiol	1.5	3.0
<b>1146</b>	<b>Camphor*</b>	<b>11.6</b>	<b>1.7</b>	1602	<i>n</i> -Hexadecane	0.4	0.9
1165	Pinocarvone	0.1	0.2	1619	<i>trans</i> -Isolongifolanone	0.1	-
1168	Borneol	2.1	0.2	<b>1633</b>	<b>Isospathulenol</b>	<b>0.4</b>	<b>4.3</b>
1176	<i>cis</i> -Pinocamphone	0.2	0.3	1649	$\beta$ -Eudesmol	0.5	0.1
<b>1180</b>	<b>Terpinen-4-ol</b>	<b>8.3</b>	<b>1.2</b>	1651	$\alpha$ -Cadinol	0.6	3.3
1193	$\alpha$ -Terpineol	0.4	0.1	1686	<i>epi</i> - $\alpha$ -Bisabolol	0.3	1.7
1198	Myrtenol	0.6	Tr.	1701	<i>n</i> -Heptadecane	0.6	-
1207	<i>n</i> -Decanal	0.1	Tr.	1722	Chamazulene	0.9	0.8
1209	<i>trans</i> -Piperitol	0.1	-	1797	<i>n</i> -Octadecane	0.1	-

RI: retention indices on HP-5 capillary column.

Tr.: trace (< 0.05%).

\*Co-injection with authentic compounds.

## EXPERIMENTAL

**Plant Material.** The flowers and leaves of *A. hyalina* were collected from Ghazvin province (northwest Iran) in May 2004 at an altitude of 1900 m. The plant was identified at the Botany Department of Isfahan University, and a voucher specimen (No. 1148) was deposited at the Herbarium of the Faculty of Pharmacy and Pharmaceutical Sciences, Isfahan University of Medical Sciences, Isfahan, Iran.

**Isolation of Essential Oil.** Plant material was hydrodistilled in a Clevenger-type apparatus for 3 h according to the method recommended in the British Pharmacopoeia [17]. The essential oil was dried over anhydrous sodium sulfate and stored in sealed vials at 4°C until analysis. The yield of oil was calculated based on dried weight of plant material.

**Gas Chromatography.** The oil was analyzed on a Perkin-Elmer gas chromatograph Model 8500, equipped with a FID detector and a BP-1 capillary column (30 m × 0.25 mm; film thickness 0.25 µm). The oven temperature was programmed from 60°–280°C at 4°C/min. The carrier gas was nitrogen with a flow rate of 2mL/min. Injector and detector temperatures were 280°C.

**GC-MS.** Gas chromatography combined with mass spectrometry was used for identification of the components detected. The analysis was performed on a Hewlett-Packard 5972A mass selective detector coupled with a Hewlett-Packard 6890 gas chromatograph, equipped with a HP-5MS capillary column (30 m × 0.25 mm; film thickness 0.25 µm) and operating under the same conditions as described above. The MS operating parameters were: ionization voltage, 70 eV; ion source temperature, 200°C.

**Identification of the Components.** Identification of components of the oils was based on GC retention indices relative to *n*-alkanes and computer matching with the Wiley 275.L library, as well as by comparison of the fragmentation patterns of the mass spectra with those reported in the literature [18, 19]. Whenever possible, the constituents were matched by co-injection with authentic compounds.

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